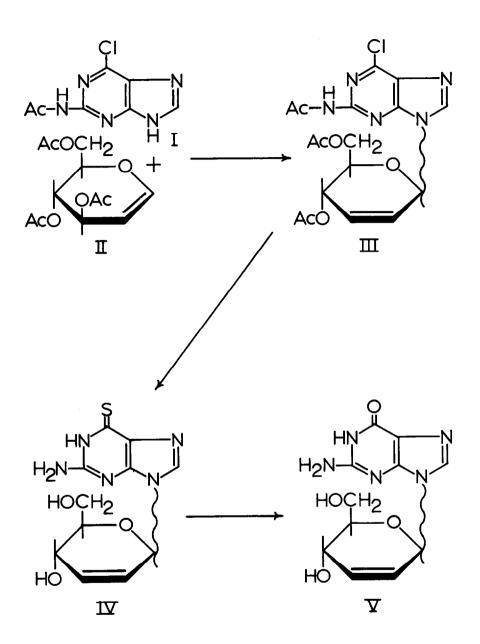
THE DIRECT UTILIZATION OF UNSATURATED SUGARS IN NUCLEOSIDE SYNTHESES. AN APPROACH TO THE PREPARATION OF ANALOGS OF BLASTICIDIN S. SYNTHESIS OF 9-(2',3'-DIDEHYDRO-2',3'-DIDEOXY-D-ERYTHRO-HEXOPYRANOSYL)GUANINE (1)

Eldon E. Leutzinger, Roland K. Robins and Leroy B. Townsend Department of Chemistry, University of Utah, Salt Lake City, Utah 84112 ('Received in USA 7 June 1968; received 'in UK for publication 27 July 1968) We wish to report a new procedure for the synthesis of a 9-pyranosylpurine possessing 2', 3'-unsaturation and a general synthetic approach to 2',3'-unsaturated pyranosyl nucleosides structurally related to Blasticidin S. This procedure has been employed in the synthesis of the guanosine analog 9-(2',3'-didehydro-2',3'-dideoxy-D-erythro-hexopyranosyl)guanine (V).

The proposal (2,3) that naturally occurring ribonucleosides might be converted into deoxynucleosides via a 2',3'-unsaturated intermediate created considerable interest which resulted in the chemical preparation of several 2',3'-unsaturated furanosyl nucleosides (4,5) of purines and pyrimidines. The isolation (6) and recent structure elucidation (7,8) of the nucleoside antibiotic blasticidin S has revealed blasticidin S to be a pyranosyl derivative of cytosine with an endocyclic double bond at the 2',3' positions. Blasticidin S has exhibited significant antifungal activity against Piricularia oryzae in rice blast disease (9), inhibition of several transplantable animal tumors (10) and more recently has shown a definite inhibition of polypeptide synthesis (11) in an <u>E</u>. <u>coli</u> strain of B cells. A mixture of 2-acetamido-6-chloropurine (I) and 3,4-6-tri-0-acetyl-D-glucal (II) was fused at 140° in the presence of a catalytic amount of trichloroacetic acid to furnish 30% yield of a white crystalline solid (III, m.p. 103-104<sup>0</sup>). The pmr spectrum of III revealed the presence of an absorption peak at  $m{\delta}$  2.5 (3 protons) which was assigned to the 2-acetamido group on the basis of pmr spectrum analysis of I. The absence of two additional protons at 52.5 indicated that the nucleoside material was not a 2'-deoxypyranoside (12). There was observed two singlet absorptions (3 protons each) in the \$2.05-2.15 region which corresponded to acetyl groups on the carbohydrate moiety. This represented the loss of one acetyl group from II via an allylic expulsion of the C-3 acetate group (13) during nucleoside formation (14). The remaining absorption peaks in the pmr spectrum (for the carbohydrate moiety) were partially assigned utilizing a comparison of the chemical shift data [5 6.55 (H-1', multiplet), 5 6.1-6.3 (H-2' and H-3', multiplet), 5 5.48 (one proton, multiplet) and 5 4.1-4.4 (3 protons, multiplet)]

4475



observed for III and the chemical shift data reported (15,16) for certain 4,6-di-0-acetyl-2,3didehydro-2,3-dideoxy-D-erythro-hexosides. The multiplet at 5 5.48 is presumably the H-4' proton and the multiplet at 6 4.1-4.4 can be assigned to the H-5', H- $6_{ax}$  and H- $6_{eq}$  protons. The nucleoside product obtained from the fusion reaction was assigned the structure 2-acetamido-6-chloro-9-(4',6'-di-O-acetyl-2',3'-didehydro-2',3'-dideoxy-D-erythro-hexopyranosyl)purine (III) on the basis of the above pmr data and elemental analysis (17). The actual site of glycosidation was established in the next step. Treatment of III with a methanolic solution of sodium hydrosulfide resulted in a facile nucleophilic displacement of the 6-chloro group with a concomitant removal of the blocking groups to furnish a 41% yield of 2-amino-9-(2',3'-didehydro-2',3'-dideoxy-Dervthro-hexopyranosyl)purine-6-thione (IV); m.p. 187-188°. It was established that complete deacetylation had occurred by virtue of the absence of any absorption peaks in the pmr spectrum of IV in the  $\delta$  2.0-2.5 region. A comparison of the ultraviolet spectra of IV [  $j_{max}^{pH~1}$  261.5 nm, 345  $\lambda_{max}^{pH}$  11 251.5 nm; 318 nm (47500, 11000)] with the spectra of 2-amino-1-methylnm (+4600, 12000; purine-6-thione (18), 2-amino-3-methylpurine-6-thione (19), 2-amino-7-methylpurine-6-thione (20) and 2-amino-9-ethylpurine-6-thione (21) firmly established the actual site of glycosidation for IV as N-9. Thus the precursor III, is also a purine-9-glycoside. The sulfur atom at position six was exchanged for an oxygen atom by treatment of IV with hydrogen peroxide in a 20% aqueous ammonia solution to furnish a 78% yield of 2-amino-9-(2',3'-didehydro-2',3'-dideoxy-D-erythrohexopyranosyl)purin-6-one (V) [ $\int_{max}^{pH \ l}$  253 nm, 275 nm (fl1500, 8200),  $\int_{max}^{pH \ ll}$  262 nm (fl2000)]. A comparison of the pmr spectrum of V with the pmr spectrum of IV revealed virtually no change in that region of the spectrum attributed to the carbohydrate moiety (\$3.0-7.0). Retention of the absorption peaks assigned to the vinylic protons (S 6.1-6.3, multiplet) firmly established that the oxidation step had occurred without affecting the 2',3'-endocyclic double bond. Extension of the fusion procedure utilizing unsaturated sugars in novel nucleoside syntheses is an area under continuing investigation in our laboratory.

## REFERENCES

- 1. This work was supported by Contract PH 43-65-1041 with the Cancer Chemotherapy National Service Center, National Cancer Institute, National Institutes of Health, U. S. Public Health Service.
- 2. P. Reichardt, J. Biol. Chem., 237, 3513 (1962).
- 3. J. P. H. Verheyden and J. G. Moffatt, J. Am. Chem. Soc., 86, 1236 (1964).
- 4. J. R. McCarthy, Jr., M. J. Robins, L. B. Townsend and R. K. Robins, <u>J. Am. Chem. Soc.</u>, <u>88</u>, 1549 (1966).

- 5. J. P. Horwitz, J. Chua and M. Noel, Tetrahedron Letters, 1343 (1966).
- S. Takeuchi, K. Hirayama, K. Ueda, H. Sakai and H. Yonehara, J. <u>Antibiot</u>. (Japan), <u>11A</u>, 1 (1958).
- 7. J. J. Fox and K. A. Watanabe, Tetrahedron Letters, 897 (1966).
- 8. H. Yonehara and N. Otake, <u>Tetrahedron</u> Letters, 3785 (1966).
- F. Misato, I. Ishii, M. Asahawa, Y. Okimoto and K. Fukunaga, <u>Ann. Phytopath. Soc.</u> (Japan), <u>24</u>, 302 (1959).
- 10. N. Tanaka, Y. Sakagami, H. Yamaki and H. Umezawa, J. Antibiot. (Japan), 14A, 123 (1961).
- 11. H. Yamaguchi and N. Tanaka, J. Biochem., 60, 632 (1966).
- E. E. Leutzinger, W. A. Bowles, R. K. Robins and L. B. Townsend, J. <u>Am. Chem. Soc.</u>, <u>90</u>, 127 (1968).
- 13. R. J. Ferrier, J. Chem. Soc., 5443 (1964).
- 14. W. A. Bowles and R. K. Robins, J. Am. Chem. Soc., 86, 1252 (1964). See Ph.D. Thesis, W. A. Bowles, Arizona State University, 1964.
- 15. D. M. Ciment and R. J. Ferrier, J. Chem. Soc., 441 (1966).
- 16. R. J. Ferrier, W. G. Overend and G. H. Sankey, J. Chem. Soc., 2830 (1965).
- 17. Satisfactory analytical data were obtained for all compounds which were also shown to be homogeneous by thin layer chromatography.
- 18. C. W. Noell, D. W. Smith and R. K. Robins, J. Med. Pharm. Chem., 5, 996 (1962).
- 19. L. B. Townsend and R. K. Robins, J. Amer. Chem. Soc., 84, 3008 (1962).
- 20. R. N. Prasad and R. K. Robins, J. Amer. Chem. Soc., 79, 6401 (1957).
- 21. C. W. Noell and R. K. Robins, J. Med. Pharm. Chem., 5, 558 (1962).